Preliminary communication

The antithrombin-binding sequence of heparin studied by n.m.r. spectroscopy

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Heparin prevents the coagulation of blood by binding to, and thus activating, antithrombin¹. Only a fraction of the molecules in heparin preparations bind with high affinity to this protein, and this fraction accounts for most of the anticoagulant activity²⁻⁴.

In order to define the antithrombin-binding sequence of the active polysaccharide-molecules, oligosaccharides with high affinity for antithrombin were isolated from partially depolymerised (nitrite-degraded) heparin by affinity chromatography on immobilised antithrombin⁵. The oligosaccharides were fractionated further according to molecular weight by gel chromatography on Sephadex G-50; the smallest, high-affinity fragment isolated was an octasaccharide⁶. Structural studies revealed a major sequence, α -L-IdAp- $(1\rightarrow 4)$ - α -D-GlcNAcp- $(1\rightarrow 4)$ - β -D-GlcAp- $(1\rightarrow 4)$ - α -D-GlcNSO $_3$ p- $(1\rightarrow 4)$ - α -L-IdAp- $(1\rightarrow 4)$ - α -D-GlcNSO $_3$ p- $(1\rightarrow 4)$ - α -L-IdAp- $(1\rightarrow 4)$ - α -D-GlcNSO $_3$ p- $(1\rightarrow 4)$ - α -L-IdAp- $(1\rightarrow 4)$ - α -D-GlcNSO $_3$ p- $(1\rightarrow 4)$ - α -D-GlcNSO $_3$ p- $(1\rightarrow 4)$ - α -L-IdAp- $(1\rightarrow 4)$ - $(1\rightarrow$

Earlier studies by ${}^{1}H$ - and ${}^{13}C$ -n.m.r. spectroscopy demonstrated that heparins from different sources consist mainly of $(1\rightarrow 4)$ -linked α -L-idopyranosyluronic acid 2-sulfate (5) and 2-deoxy-2-sulfamino- α -D-glucopyranosyl 6-sulfate (1) residues 8 —10. Small

amounts of β -D-glucopyranosyluronic acid (6) residues, non-sulfated α -L-idopyranosyluronic acid (4) residues, and 2-acetamido-2-deoxy-D-glucosyl (3) residues have also been demonstrated8. The 13C-n.m.r. spectrum obtained from native, pig-mucosal heparin (Fig. 1, B) is in good agreement with these previous results. By contrast, the spectrum produced by the high-affinity decasaccharide (A), isolated from the same heparin preparation, is markedly different (Fig. 1, A) and resembles that of the oligosaccharide isolated by fractionation of commercial (Choay), pig-mucosal heparin 11. Signals assigned to residues I and 5, the major components of the intact polysaccharide chain, are clearly seen but, in addition, there are prominent signals ascribed to residues 3, 4, and 6, which are all minor constituents of the heparin molecule. Assignments of most of the signals in the 13 C-n.m.r. spectrum of A are made by reference to earlier studies $^{8-11}$. Thus, the peaks at δ 95.35, 53.69, and 22.06 (not included in Fig. 1) are assigned to C-1, C-2, and the N-acetyl group of 3, respectively. The signals for C-2 of 1 units appear at δ 57.34 and 57.65, reflecting attachment to different uronic acid residues (one is probably the non-sulfated L-iduronic acid which occurs at the non-reducing end of the octasaccharide sequence shown above). The signal for C-1 of 2-sulfated L-iduronic acid (5) appears at δ 99.23, the anomeric carbon atom of unsubstituted L-iduronic acid (4) appears at δ 100.96, and that of D-glucuronic acid (6) at δ 101.89. The carbonyl carbon atoms from carboxyl and N-acetyl groups appear at δ 174.6-175.1 (not included in Fig. 1). The signal at δ 56.54 is attributed to C-2 of the unique, 3-sulfated residue 2. This assignment is made by comparison with the corresponding signal from 1, and adjustment for the β shift effect on C-2 of an O-sulfate group at C-3. The shift effect was determined by comparing the spectra of methyl 2-amino-2-deoxy-α-D-glucopyranoside (δ 96.32, C-1; 54.28, C-2; 70.15, C-3; 69.78, C-4; 72.17, C-5; and 60.62, C-6) and its 3-sulfate (8) 96.29, C-1; 53.33, C-2; 77.91, C-3; 67.96, C-4; 71.71, C-5; and 66.44, C-6). Further, in the ¹H-n.m.r. spectrum of A, H-3 of 2 exhibits a shift of δ 4.39 which is ~0.7 p.p.m. downfield of the corresponding signal for 1.

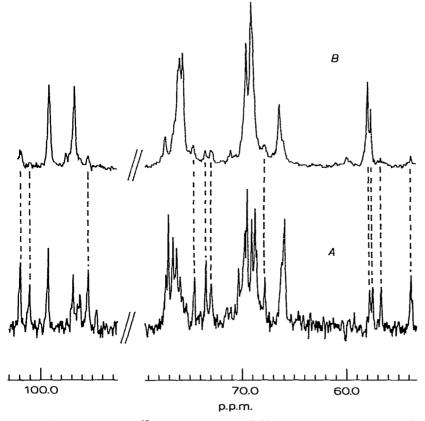


Fig. 1. Expansion of the 13 C-n.m.r. spectra (67.89 MHz; broad-band decoupled; reference, internal acetone, δ 30.5; 30°) of the antithrombin-binding oligosaccharide of heparin (A) and native heparin (B).

These results are in good agreement with the structure proposed for the antithrombin-binding sequence in heparin⁷. In particular, the postulated position of the unique sulfate group, at position 3 of a 2-amino-2-deoxy-D-glucosyl residue⁷, is confirmed.

Most of the minor signals from the intact heparin chains (Fig. 1, B) correspond to major signals of the high-affinity decasaccharide (Fig. 1, A). The antithrombin-binding sequence thus shows a high proportion of sugar residues that are poorly represented in the heparin molecule as a whole (residues 2, 3, 4, and 6). The polymer regions outside the binding sequence should be largely composed of repeating disaccharide-units (residues 1 and 5).

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REFERENCES

- 1 R. D. Rosenberg, Fed. Proc., Fed. Am. Soc. Exp. Biol., 36 (1977) 10-18.
- 2 L. H. Lam, J. E. Silbert, and R. D. Rosenberg, Biochem. Biophys. Res. Commun., 69 (1976) 570-577.
- 3 M. Höök, J. Björk, J. Hopwood, and U. Lindahl, FEBS Lett., 66 (1976) 90-93.
- 4 L. O. Andersson, T. W. Barrowcliffe, E. Holmer, E. A. Johnson, and G. E. C. Sims, *Thromb. Res.* 9 (1976) 575-583.
- 5 U. Lindahl, G. Bäckström, M. Höök, L. Thunberg, L. Å. Fransson, and A. Linker, Proc. Natl. Acad. Sci. U.S.A., 76 (1979) 3198-3202.
- 6 L. Thunberg, G. Bäckström, H. Grundberg, J. Riesenfeld, and U. Lindahl, unpublished results.
- 7 U. Lindahl, G. Bäckström, L. Thunberg, and I. G. Leder, Proc. Natl. Acad. Sci. U.S.A., in press.
- 8 A. S. Perlin, N. M. K. Ng Ying Kin, S. S. Bhattacharjee, and L. F. Johnson, Can. J. Chem., 50 (1972) 2437-2441.
- 9 A. S. Perlin, B. Casu, G. R. Sanderson, and L. F. Johnson, Can. J. Chem., 48 (1970) 2260-2268
- 10 G. Gatti, B. Casu, and A. S. Perlin, Biochem. Biophys. Res. Commun., 85 (1978) 14-20.
- 11 J. Choay, J. C. Lormeau, M. Petitou, P. Sinay, B. Casu, P. Oreste, G. Torri, and G. Gatti, Thromb. Res., 18 (1980) 573-578.
- 12 I. G. Leder, Biochem. Biophys. Res. Commun., 94 (1980) 1183-1189.